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# In vitro TOXICITY OF DECABROMODIPHENYL ETHANE (DBDPE)

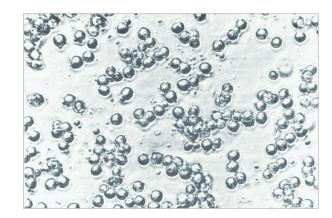
#### Introduction

Decabromodiphenyl ethane (DBDPE) came into the market in the early 1990s as an alternative to decabromodiphenyl ether (BDE-209), which was assumed not to be bioavailable due to its high molecular weight of 971 g/mol. However, the BDE-209 has been detected in humans as well in the environment. DBDPE has been used as an additive flame retardant in different types of plastics and textiles, and it is predicted to become one of the major flame retardants used by the thermoplastics industry. There is evidence of its existence in aquatic food web but amount of data concerning the toxicity of DBDPE is limited.

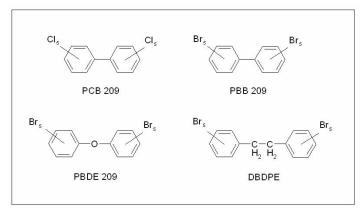
## Materials and Methods

Freshly isolated hepatocytes from hatchery-reared brown trout (*Salmo trutta*) males were used to test the bioaccumulation, detoxification metabolism and estrogenic activity of DBDPE. Ethoxyresorufin-O-deethylase (EROD) and uridinediphosphoglucuronyltransferase (UDPGT) activities were used to assess the detoxification activity. Cell vitellogenin induction was the variable for the estrogenic activity.

The hepatocyte cell cultere samples were extracted with dichloromethane (DCM). The solvent was changed to isooctane and the extract was treated with concentrated sulphuric acid. The quantitation was done with <sup>13</sup>C-labelled DBDPE as an internal standard. The analyses were conducted on Varian CP-3800 GC coupled to Varian 1200L triple quadrupole mass spectrometer. Short analytical column (10m x 0.53mm) was operated at diminished pressure (Rapid-MS<sup>TM</sup>). Mass spectrometer was operated at El/SIM mode. The most abundant fragment and the molecular ion was monitored (m/z 971 [M+10]<sup>+</sup> and 485 [C<sub>6</sub>Br<sub>5</sub>CH<sub>2</sub>]<sup>+</sup>).



Pic. 1. Trout hepatocytes after 2h in culture.\*\*

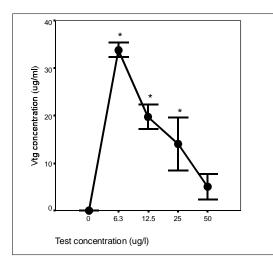


Pic. 2. Molecular structures of PCB-209, PBB-209, PBDE-209 and DBDPE

### Results

DBDPE was detected from the hepatocytes at concentrations corresponding accumulation of 42% (26 ng/18 milj cells) at test concentration of 6.25  $\mu$ g/l with decrease to 17% (84 ng/18 milj. cells) at test concentration of 50  $\mu$ g/l. DBDPE was not detected from the cell culture blank or from the procedual blank.

DBDPE caused a significant vitellogenin induction in the separated hepatocytes already at the concentration of  $6.3 \mu g/l$  (Fig.1.). Inhibition was observed at higher test concentrations. Hepatocyte EROD activity (Fig.2.) followed similar courses with that of the vitellogenin. The activity increased significantly up to the concentration of  $12.5 \mu g/l$ . After that DBDPE started to inhibit the enzyme activity so that at the concentration of  $25 \mu g/l$  the cell EROD activity was decreased to zero. What comes to the cell UDPGT activity (Fig.3.) the trend was increasing till the highest test concentration,  $50 \mu g/l$ .



ig. 1. Vitellogenic response (mean ± SD) of

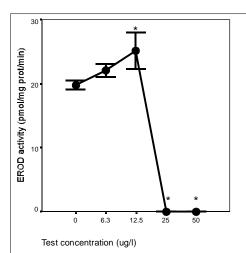


Fig. 2. Hepatocyte EROD activity. Asterisks

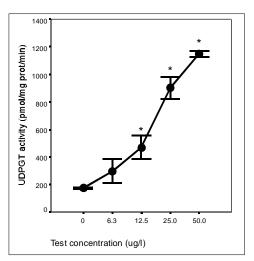


Fig. 3. Hepatocyte UDPGT activity. Asterisks

hepatocytes to DBDPE. Asterisks denote the significant difference from the control (p<0.05).

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#### Conclusions

Even though the assumption has been that DBDPE is not bioavailable it was observed that DBDPE does accumulated into the hepatocytes. According to the observed changes in the hepatocyte detoxification activities it can be stated that there is metabolism of DBDPE in the brown trout liver cells. The toxic effects of DBDPE and the effective concentration levels has to be investigated. Also the occurrence of DBDPE in the environment needs to be further studied.

#### References

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